

MICROBIAL DIVERSITY IN RIVER WATER AND SEDIMENT FROM SULINA AREA

**CÎRSTEA Doina Maria, CIOBOIU Olivia, DUMITRACHE Alina Cristina,
ȘTEFĂNESCU Mugur Cristian**

Abstract. Research on the bacterial diversity of the Danube waters from the flowing or stagnant area, somewhat adjacent to Sulina branch and port, is a study case integrated with traditional ecological research, contributing to a more complex characterization of the permanent or accidental populations from these areas. The differences or similarities with respect to the benthic microbiota and the one present in the water mass, as well as the abiotic and biotic characteristics of the delta channels and lakes, correlated with the seasonal climatic variations or generated by the extreme or deficient pluviometric regime, constitute extremely complex and interesting study material that will be able to supplement the results of the research undertaken so far.

Keywords: bacterial diversity, benthic microbiota, sediments samples.

Abstract. Diversitatea microbială a apei și a sedimentelor din zona Sulina. Cercetările privind diversitatea bacteriană a apelor Dunării, din zona curgătoare sau stagnantă, limitrofă într-o oarecare măsură brațului și portului Sulina, constituie un caz de studiu, integrat cercetărilor ecologice deja cu tradiție, contribuind la o mai complexă caracterizare a populațiilor de organisme, permanente sau accidentale din aceste areale. Diferențele sau similitudinile în ceea ce privește microbiota bentonică și cea prezentă în masa apei, cât și caracteristicile abiotice și biotice ale apelor lacurilor de deltă și ale canalelor, corelate cu variațiile climatic sezoniere sau generate de regimul pluvial extrem sau deficitar, constituie un material de studiu extrem de complex și de interesant care va putea să completeze rezultatele cercetărilor întreprinse până în prezent.

Cuvinte cheie: diversitate bacteriană, microbiotă bentonică, probe de sediment.

INTRODUCTION

The studies undertaken to fully characterize the biodiversity of the Danube River, supplement the projects of mapping and distribution of rare and protected species of plants and animals, in the enhanced interest of protecting the environment, not only in the Danube Delta Biosphere Reserve, but anywhere where accurate knowledge of fish, ornithological, botanical and microbial groups is required (KIRSCHNER et al., 2009). In this respect, we are working on the application of the most appropriate measures required by specialists from very diverse but related fields, thus justifying the interest of carrying out studies on the microbiota in the delta area, adjacent to Sulina Ecological Station, a research point belonging to the Institute of Biology Bucharest, under the aegis of the Romanian Academy.

The present paper is part of a series of three studies, based on the analysis of the samples taken in September 2016 from the Delta lakes Puiu, Rosu and Rosulet, as well as from the related linking channels. The papers are subject to presentations at international scientific events organized by the University of Pitesti and by the County Museum of Oltenia during 2017. Thus, at the Scientific Symposium Current Trends in Natural Sciences organized in Pitești in April 2017, the paper "Highlighting of some physiological bacterial groups isolated from the Danube Delta Biosphere Reserve, Rosu, Roșuț and Puiu lakes" was presented, which signals of the main physiological groups of microorganisms present in the water and benthos samples taken from several delta points where previous environmental analyses were performed.

In the other two papers brought to the attention of specialists and of those interested in the microbial diversity of the Danube Delta, we present a physico-chemical characterization of both water and benthic soil samples on the one hand, and an interpretation of the presence of certain bacterial physiological groups (PĂCEȘILĂ et al., 2008), thus making a comparison between the microbiota present in the water mass and the one identified in the sediment layer.

At the same time, by isolating and testing bacterial populations from the same analyzed samples, selections of bacterial strains with biotechnological potential were made. These researches on the bacterial cell metabolism and the possible uses of the obtained bioproducts are the subject of the third scientific communication that will be presented this autumn at the international scientific manifestation organized by the Oltenia Museum, Craiova.

MATERIAL AND METHODS

Field sampling. Water samples were collected in September 2016. The analyzed Danube water sampling points were located on Busurca Channel (M3), Împușita Channel, at the confluence with Rosuleț Lake (M4), in Roșu Lake (M6), at the mouth of Roșu Lake (M8), and in Mândra backwater (M9).

For the sediment samples, the chosen sampling sites were Împușita Channel at the confluence with Roșuleț Lake (M4), Roșu Lake (M6), Musura Gulf (3) and Sulina harbor area.

The samples consisted of 500 mL of column water from each selected station point; some water was filtered through GF / F filters (65 μm) for dissolved forms of nutrients and preserved for further analyses.

Sediment samples: surface sediments samples were also sampled and transported in plastic bags to the lab and preserved at -20 °C until extraction and analysis.

Measurement of physical parameters. Depth and transparency were determined using Secchi disc, the redox potential, pH, temperature, conductivity and salinity were measured in the field with a multiparameter WTW 340i, Germany.

Water chemistry. Nutrients were determined spectrophotometrically following a modified Berthelot method for N-NH₄ (KROM, 1980), Griess-Ilosvay modified method for N-NO₂ (KEENEY & NELSON, 1982; TARTARI & MOSELLO, 1997) for N-NO₃, P-PO₄ and TP.

Selective growing media. In order to highlight the main physiological groups of bacteria present in the analyzed water and sediment samples, the following selective cultivation media were used: VL medium (BEERENS, 1954), for anaerobic heterotrophic bacteria, Postgate for reducing sulphate bacteria, Nutrient Broth for aerobic heterotrophic facultative anaerobic bacteria, medium with starch for amylolytic bacteria, Vinogradsky for ferrobacteria, medium for nitrite bacteria, Giltay medium for denitrifying bacteria and 9K medium for iron oxidizing chemolithotrophic bacteria. Serial dilutions were grown on the selective media in triplicates (three repetitions) and incubated at 28 °C for varying lengths of time. The bacterial growth was assessed according to the McReady comparative method.

RESULTS AND DISCUSSIONS

A first step was to establish the physico-chemical parameters of both the water in the sampling points and the collected sediment.

As it can be seen in Tables 1 and 2, parameters such as temperature, conductivity, transparency, pH, NO₂, NO₃, etc. have been measured.

In the case of water set parameters, it can be observed that there are no major fluctuations between the five sampling points. The measured parameters fall within the set limits for river water. Table 1 presents the physico-chemical analysis of the collected Danube water samples.

Table 1. Physico-chemical analysis of water.

Parameter/ station	M3 (Busurca Channel)	M4 (Împuțita Channel in Roșuleț Lake)	M6 (Roșu Lake)	M8 (channel entrance in Roșu Lake)	M9 (Mândra backwater)
Depth (m)	2	1.1	2.7	1.6	1.7
Transparency (m)	0.4	1.1	0.4	0.4	0.4
T/A	0.2	1.0	0.15	0.25	0.24
Temperature (°C)	17.5	16.2	16.9	17.1	16.9
pH	7.91	8.114	8.96	8.491	8.602
Conductivity (ms/cm)	421	545	385	386	441
Salinity	0.1	0.2	0.1	0.1	0.1
Redox	-56.9	-56.6	-110.5	-90.5	-92.6
NH ₄ (mgN/L)	0.007	0	0.024	0.088	0.054
NO ₂ (mg/L)	0.003	0.001	0.001	0.009	0.001
NO ₃ (mg/L)	0.33	0.25	0.30	0.15	0.53
DIN (mgN/L)	0.336	0.252	0.326	0.247	0.583
PO ₄ (µg/L)	21.25	10	26.25	32.5	32.5
Whole P (µg/L)	30	56.25	68.75	82.5	70

The physico-chemical parameters of the sediment were assessed using the same method used to establish the physico-chemical parameters of the water. As it can be seen in Table 2, the sediment samples originated from Împuțita Channel and Roșu Lake. Water samples were also collected from the same points. Thus, the temperature, redox and pH parameters were similar for water and sediment samples, while parameters such as salinity, conductivity, increased, showing a double value in the sediment samples as compared to those of the water at the same sampling point. This is due to a higher degree of mineralization in the sediment, which led to increased concentrations of ammonium, nitrite, nitrate, orthophosphate ions.

Table 2. Physico-chemical analysis of the sediment.

Parameter/station	M4 (Împuțita Channel in Roșuleț Lake)	M6 (Roșu Lake)	Musura Channel	Sulina Harbour
Depth (m)	1.1	2.7	1.6	1.0
pH	8.149	8.639		
Redox	-64.5	-94.5		
Temperature (°C)	19.3	19		
Salinity	0.4	0.4		
Conductivity (ms/cm)	940	855		
NH ₄ mg/g s.u	17.76	22.48	11.17	33.96
NO ₂ mg/g s.u	0.623	0.516	0.073	0.065
NO ₃ mg/g s.u	8.38	7.28	1.45	1.72
PO ₄ µg/g s.u	3488.25	5892.57	1642.47	1411.17

In the case of sediment samples taken from Musura Channel and Sulina Harbor, the ammonium, nitrite, nitrate, orthophosphate ions quantities were found to be lower in comparison to the sediment samples M4, M6, due to the presence of denitrifying bacteria present in large number in the sediment, depleting the nitrogen compounds from the soil by reducing the nitrates and nitrites. Both sediment and water samples were analyzed from the microbiological point of view, by seeding on specific media of different physiological groups. In this study, we focused on signaling the presence of the dominant physiological groups of microorganisms in both the analyzed water and sediment samples (Tables 3; 4).

Table 3. Limit value number of bacteria/ml water sample.

	M3 (Busurca Channel)	M4 (Împușita Channel in Roșuleț Lake)	M6 (Roșu Lake)	M8 (channel entrance in Roșu Lake)	M9 (Mândra backwater)
No. of cell/ml	11 x 10⁵ sulphate reducing bacteria – 15 x 10⁸ iron bacteria	3 x 10³ heterotrophic anaerobic bacteria – 11 x 10⁶ iron bacteria	9.5 x 10³ sulphate reducing bacteria; heterotrophic aerobic and facultative anaerobe bacteria; amylolytic bacteria – 11 x 10⁴ heterotrophic anaerobic bacteria	0.4 x 10³ heterotrophic anaerobic bacteria – 11 x 10⁷ nitrite bacteria	0.4 x 10² iron bacteria – 11 x 10⁷ nitrite bacteria

Table 4. Limit value bacterial count/ml sediment sample.

	M4 (Împușita Channel in Roșuleț Lake)	M6 (Roșu Lake)	Musura Channel	Sulina Harbour
No. of cell/ml	1.5 x 10 Iron-oxidizing chemolithotrophic bacteria – 4.5 x 10⁷ Heterotrophic anaerobic bacteria	4.5 x 10 Iron-oxidizing chemolithotrophic bacteria – 11 x 10⁹ heterotrophic anaerobic and facultative anaerobic bacteria	9.5 x 10 Iron-oxidizing chemolithotrophic bacteria – 11 x 10⁹ Denitrifying bacteria	2.5 x 10 Iron-oxidizing chemolithotrophic bacteria – 14 x 10⁸ Heterotrophic anaerobic bacteria

It should be noted that in the water samples from the Busurca and Împușita Channels ferrobacteria are predominant, while in Roșu Lake and in Mândra backwater the nitrite bacteria are preponderant. In the analyzed sediment samples, the bacteria belonging to the anaerobic heterotrophic group predominate, and in Musura Bay the denitrifying bacteria were present.

The comparison between the two sets of samples, water and sediment, reveals that in all four sediment samples there are minimal and relatively constant concentrations of iron-oxidizing chemolithotrophic bacteria. It can be concluded that the sedimentary benthic layer, due to the presence of aerobic facultative anaerobic bacteria, is microbial-active.

The circulating water mass, with a depth of 1.0 to 2.7 meters, varies depending on the Danube water shares at different time intervals, as well as the degree of transparency. It is necessary to periodically analyze the same sampling points in order to observe any variations of the microbiota.

CONCLUSIONS

The analyzed samples presented a wide variety of physiological groups of the microorganisms.

Ferrobacteria were predominantly present in the mass of water coming from channels with a relative circulating flow, while in the lakes with stagnant water volume there are increased amounts of nitrite bacteria.

Sediment samples taken from depths of 1 down to 2.7 meters show normal benthic microbial activity.

ACKNOWLEDGEMENTS

The study was funded by the project no. RO1567-IBB05/2016 of the Institute of Biology Bucharest of the Romanian Academy.

REFERENCES

BEERENS H. 1954. Amelioration des techniques d'etude et d'identification des bacteries anaerobies. *Annales de Institut Pasteur de Lille*. Edit. Masson & Cie. Paris. **6**: 36-52.

- KEENEY D. R. & NELSON D. W. 1982. Nitrogen - Inorganic forms. In: *Methods of soil analysis*, Page A. L., Miller R. H., Keeney D. R. (Eds.). **2**(2). *Agronomy Monograph*. ASA and SSSA. Madison. **9**: 643-698.
- KIRSCHNER A. K. T., KAVKA G. G., VELIMIROV B., MACH R. L. SOMMER R., FARNLEITNER A. H. 2009. Microbiological water quality along the Danube River: Integrating data from two whole-river surveys and a transnational monitoring network. *Water Research*. Springer. Berlin. **43**(15): 3673-3684.
- KROM M. D. 1980. Spectrophotometric determination of ammonia: a study of a modified Berthelot reaction using salicylate and dichloroisocyanurate. *Analyst. Journal of Chemistry*. Springer. Stuttgart. **19**(105): 305-316.
- TARTARI G. & MOSELLO R. 1997. *Metodologie analitiche e controlli di qualità nel laboratorio chimico dell'Istituto Italiano di Idrobiologia. Documenta dell'Istituto Italiano di Idrobiologia*. Consiglio Nazionale delle Ricerche. Universitaria Publisher. Verbania. **60**. 160 pp.
- PĂCEȘILĂ I., IONICĂ DOINA, COMAN A. 2008. Evaluation of metabolic intensity of benthic microbial communities from Musura Bay (Black Sea) using dehydrogenase activity method. *Proceedings of the 37th IAD Conference*. Lymnological Reports. Chișinău: 194-198.

Cîrstea Doina Maria, Dumitrache Alina Cristina, Ștefănescu Mugur Cristian

Institute of Biology Bucharest of the Romanian Academy, Spl. Independentei no. 296, sect. 6, 060031, Bucharest, Romania.
E-mails: doina.cirstea@ibiol.ro; maria_2na@yahoo.com; mugur.stefanescu@ibiol.ro

Cioboiu Olivia

The Oltenia Museum, Craiova, Str. Popa Șapcă, No. 8, 200422, Craiova, Romania.
E-mail: oliviacioboiu@gmail.com; cioboiu.olivia@yahoo.com

Received: March 30, 2017
Accepted: July 7, 2017